

Original Article

Prevalence of Listeria Monocytogenes in Milk

Prevalence of *Listeria Monocytogenes* in Raw Milk of the Healthy Cattle in Lorestan Province (Iran) by PCR

Ebadifar Amir Mohamad , Jaydari Amin*, Shams Nemat , Rahimi Heidar

Department of Pathobiology, Faculty of Veterinary Medicine, Lorestan University, Khorramabad, Iran

Received: 29 Jan 2022 Accepted: 16 Feb 2022

Abstract

Background & Objective: The prevalence of *Listeria monocytogenes (L. monocytogene)* in milk and dairy products, the high complications and damage associated with zoonotic Listeriosis have led to this bacterium being considered as a risky infection for public health.

Materials & Methods: In the present descriptive and cross-sectional study, to investigate the prevalence of *L. monocy-togene*, 100 samples of raw milk obtained from industrial farms in Lorestan province in winter and spring of 2019-2020 were collected and *16SrRNA* and *hlyA* genes were detected using PCR reaction to identify *Listeria* genus and *L. mono-cytogene* species, respectively.

Results: Based on the observed results, three samples (3%) were positive for *Listeria* genus, while no report of *L. mono-cytogens* species was observed.

Conclusion: The results of this study did not show evidence of the presence of *L. monocytogenes* in raw cow milk, which indicates the sanitation and desirable nutritional conditions in industrial farms. Therefore, due to the presence of *Listeria* (other species of *Listeria*) in raw milk, if you consume raw and unpasteurized milk, the risk of human infection with listeriosis should not be ignored.

Keywords: Listeria monocytogenes, Milk, Zoonosis, Cattle, Lorestan, PCR

Introduction

jabs.fums.ac.ir

L. monocytogene is one of the most important bacterial infections transmitted through raw milk and dairy products that causes listeriosis in humans and animals (1). According to the Centers for Disease Control and Prevention (CDC),

*Corresponding Author: Jaydari Amin, Department of Pathobiology, Faculty of Veterinary Medicine, Lorestan University, Khorramabad, Iran Email: jaydari.a@lu.ac.ir https://orcid.org/0000-0002-5880-5229 about 1,600 people are diagnosed with listeriosis and about 260 die each year (2). Due to the ability of this microorganism to grow at low temperatures and its transmission through food and dairy, in countries with low levels of health standards, high reports of listeriosis have been recorded (1). Intermittent excretion of L. monocytogenes in raw milk of ruminants has been reported without clinical signs of listeriosis (3, 4). Therefore, diagnosis of infected animals is

Ebadifar Amir Mohammad:https://orcid.org/ 0000-0002-4235 0768 Nemat Shams: https://orcid.org/0000-0001-8614-3720

Heidar Rahimi: https://orcid.org/0000-0003-1590-2528

Downloaded from jabs.fums.ac.ir on 2025-08-31]

Ebadifar A, et al.

essential to prevent the spread of listeriosis (5).

Using the common and valid PCR molecular detection method to identify L. monocytogenes in raw milk is more favorable compared to isolation and detection of bacteria by culture and biochemical methods, due to the time consuming nature of these methods and the increasing use of antibiotics in the diet of livestock and the probability of reporting false results. Listerio lysine O (LLO) is a secretory protein and a major causative agent of L. monocytogenes that is present in all pathogenic L. monocytogenes and is encoded by the hlyA gene, so its detection in food is an indicator of the presence of L. monocytogenes (6, 7). There are few studies related to the prevalence of L. monocytogenes in cow's milk and no such study has been performed on livestock in Lorestan province (Iran). Therefore, considering the importance of cow's milk as the main source of dairy production and the important role of milk in the possible transmission of infection to humans, in the present study, PCR method was used to directly detect L. monocytogenes to investigate the prevalence of this bacterium in cow's milk in Lorestan province.

Materials & Methods

Milk sampling

Milk samples were randomly collected from 100 cows from industrial farms in Lorestan province (Iran) from September 2019 to March 2020. 50 milliliters of each sample was gathered in tubes under aseptic conditions and transferred to laboratory on an icebox. The samples were stored at -20 °C for further studies.

DNA extraction

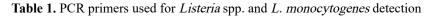
Frozen milk samples were thawed at room temperature and 10 mL of each sample was transferred to a 10 mL tube. The tubes were centrifuged at 13,000 × g for 10 min. Then, using a sterile swab, the top layer of fat was removed and the liquid was poured around. Next, 200 μ L of precipitate were transferred to a new tube for DNA extraction using DNA Purification Mini Kit (GeneAll, Korea) according to the manufacturer instructions. The quality and quantity of extracted DNA was assessed by 1% agarose gel and Nanodrop spectrophotometer, respectively (Thermo Scientific, Waltham, USA). Finally, the extracted DNA was stored at -20°C for use in PCR reaction.

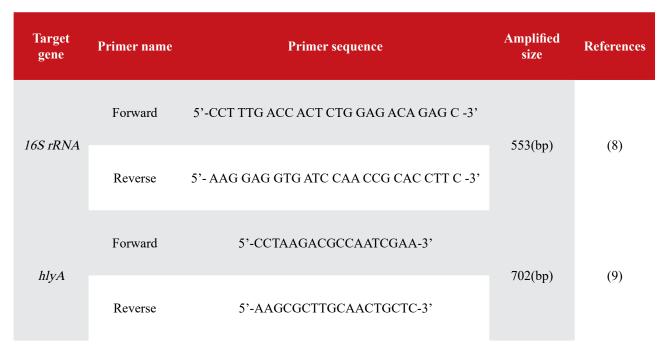
DNA amplification and detection of polymerase chain reaction (PCR) products

To identify the isolates, the PCR method was employed and specific primers of 16S rRNA (553 bp) and hlyA (702 bp) genes for identification of Listeria spp were used L. monocytogenes, respectively. PCR amplification was conducted in 25 µL total volume using 12.5 µL of 2X master mix (Ampliqon Taq DNA Polymerase Master Mix RED), 0.5 µL of each specific forward and reverse primers (10nM) (provided by Takapou Zist Company, Tehran, Iran) (Table 1), and 4 µL of the extracted DNA. Genomic DNA isolated from reference strain L. monocytogenes (PTCC 1294) (Iran Scientific and Industrial Research Center) and sterile water were considered as positive and negative controls, respectively. These amplifications were conducted by Bio-Rad thermocycler (Model T- 100, USA) under the following conditions: Program A is for Listeria spp and program B is for L. monocytogenes. A: The initial step of 94°C for 5 min was followed by 30 cycles of 94°C for 30 Second as denaturation, annealing at 60°C for 45 Second, extension at 72°C for 45 Second, and followed by final extension at 72°C for 10 min. B: The initial step of denaturation at 95°C for 5 min, followed by 30 cycles of denaturation at 95°C for 1 min, annealing at 52°C for 45 Second, extension at 72°C for 2 min, and final extension at 72°C for 8 min. The PCR products were separated in a 1.2% (w/v) agarose gel (Merck, Germany) containing 2.5 µg/mL DNA safe stain (Cinnagen, Iran). Electrophoresis was performed in 0.5x Tris/ Borate/EDTA (TBE) buffer for one hour at 100 V. The resulting PCR products were visualized under a UV transilluminator (E-Box, Iran) and the 100 bp DNA ladder (Smobio, Taiwan) plus was used as the molecular size marker.



Prevalence of *Listeria Monocytogenes* in Milk





Results

In the present study, by examining 100 samples of raw milk collected from industrial farms in Lorestan province, based on PCR results of *16s rRNA* gene, the results

showed that 3 samples (3%) were infected with *Listeria spp* (Figure 1). But, none of the samples of *L. monocytogenes* was detected by PCR results of *hlyA* gene (Figure 1).

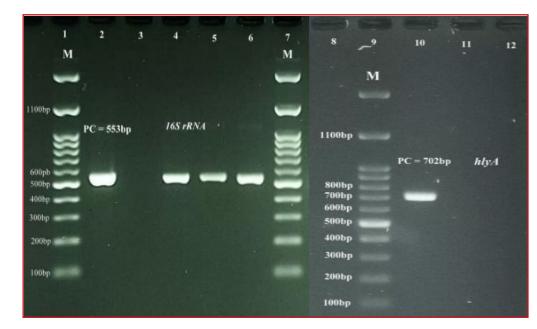


Figure1. PCR assay for the detection of *Listeria spp. and L. monocytogenes* in raw milk samples. Lane 1, 7, 9 (M) Standard DNA marker; Lane 2: Positive control for *Listeria spp*; Lane 3: Negative control; Lanes 4–6: Positive samples for *Listeria spp*; Lane 8: empty; Lane 10: Positive control for *L. monocytogenes*; Lane 11: Negative control; Lanes 12: Negative samples for *L. monocytogenes*. (PC: Positive control)

Discussion

Listeriosis is a zoonotic disease with food originality. Due to the widespread prevalence of Listeria species in the environment and the ability of this bacterium to grow at temperatures of 0 to 4 ° C and its high resistance to osmotic pressure, raw milk and contaminated dairy products are the most important route of Listeriosis transmission, so that in many studies, more emphasis has been placed on identifying listeria isolates from raw milk samples (4, 10, 11). In the present study, the prevalence of Listeria in milk samples isolated from industrial dairies in Lorestan province was determined to be 3%. While, L. monocytogenes was not detected in any of the samples. Numerous studies in different countries have examined the prevalence of L. monocytogenes in cow's raw milk. This prevalence was reported 4% In the United States (12), 2% in Turkey (13), 4% in Netherlands (14), 3.6% in Ethiopia (15), 1.7% in Italy (16), 1.1% in Brazil (17), 11% in Iraq (18) and 1.7% in Iran (19) and 1/1% in Iran (11). However, the present study is in line with the studies of Rahimi et al (11) and Mohammadi (19) which show a low prevalence of L. monocytogenes in cow's milk and indicates the low prevalence of L. monocytogenes in cattle population of Iranian livestock.

According to studies, one of the important sources of milk contamination with *Listeria isolates*, especially *L. monocytogenes*, is the feeding with low quality silage and lack of sanitation during milking (20, 21). However, in the present study, the selected cows not only had a history of feeding on silage, but during milking from the cow's udder, the nipples were disinfected with 70% alcohol. Therefore, the amount of detected infections by *Listeria* and *L. monocytogenes* in this study can confirm the observance of sanitation and nutritional conditions in the target farms.

The present study, consistent with other studies conducted in Iran, Rahimi et al. (11) and Jamali et al. (22), shows that the prevalence of *listeria* in cow's milk is lower than sheep's and goat's milk

by comparison of the prevalence of *listeria* in cow's milk with the prevalence in sheep and goat milk in these studies. Rahimi et al reported that the prevalence of *listeria* in cow, sheep and goat were 11.1%, 22.6% and 6.7%, respectively

Ebadifar A, et al.

(11). In the study by Jamali et al, these rates of prevalence were determined to be 1.1%, 6.5% and 1.7%, respectively (22). It seems that this condition can be affected by the conditions of dense husbandry.

The present study is in agreement with the study of Iran (23) and other studies conducted in the United States (24), Brazil (25 - 27) Australia (28) and Egypt (29) which have reported 0%prevalence of L. monocytogenes in raw cow milk. Although good nutrition and sanitation are effective, studies have shown that high levels of natural microbiota in raw foods can reduce the survival of pathogens such as L. monocytogenes in food (raw milk). Because pathogens need specific conditions to grow. Therefore, the presence of high levels of natural microbiota in raw milk prevents the growth of pathogens (30 - 34). Numerous studies have shown that depending on the listeria strain and the amount of its presence in milk, this bacterium can survive in the long periods of pasteurization and even freezing at very low temperatures (35-38). Thus, despite the low prevalence and concentration of pathogens, the presence of bacteria is an important risk factor for infection caused by eating contaminated food, especially when consuming unpasteurized dairy products (39).

Conclusion

Due to the importance of cow's milk in the preparation of raw dairy products such as local cheese, which is made from raw milk in most rural areas, the potential risk of *Listeria* infection in people, especially for pregnant women, the elderly and people with defective immune systems is worrying. Therefore, high-risk groups should avoid unpasteurized dairy products. Also, due to enzymes secreted such as lipase and



proteinase from *Listeria* bacteria in sterilized milk, it is very important to observe hygienic standards, identify and treat infected animals.

Conflict of interest

The authors declare that no conflict of interest exists.

Acknowledgement

The authors would like to thank Mr. Ali Karimpor and Veterinary personnel of Noor Abad and Kuhdasht Cities for their aid in sample collection. This study is the result of Mr. Amir Mohammad Ebadifar's master thesis No. 2754786. From Lorestan University

References

1. Sarfraz M, Ashraf Y, Ashraf S. A Review: Prevalence and antimicrobial susceptibility profile of listeria species in milk products. Matrix Sci Medica. 2017; 1: 3–9.

2. Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases (NCEZID), Division of Foodborne, Waterborne, and Environmental Diseases (DFWED). Access Date: 8.03. 2022. https://www.cdc.gov/listeria/prevention.html.

3. Conly J, Johnston B. Listeria: A persistent food-borne pathogen. J Can des Mal Infect la Microbiol Medicale. 2008; 19: 327–328. doi:10.1155/2008/702565.

4. Schoder D, Melzner D, Schmalwieser A, Zangana A, Winter P, Wagner M. Important vectors for Listeria monocytogenes transmission at farm dairies manufacturing fresh sheep and goat cheese from raw milk. J Food Prot. 2011; 74: 919–924.

5. Gronstol H. Listeriosis in sheep Listeria monocytogenes excretion and immunological state in healthy sheep. Acta Vet Scand. 1979; 20: 168–79.

6. Wesely IV, Harmon KM, Dickson JS, Schwartz AR. Application of a Multiplex Polymerase Chain Reaction Assay for the Simultaneous Confirmation of Listeria monocytogenes and Other Listeria Species in Turkey Sample Surveillance†. J Food Prot. 2002;65, 780–785. doi:10.4315/0362-028X-65.5.780.

7. Choi WS, Hong CH. Rapid enumeration of Listeria monocytogenes in milk using competitive PCR. Int J Food Microbiol. 2003; 84: 79–85. doi:10.1016/S0168-1605(02)00401-4.

8. Lantz PG, Hahn-Hägerdal B, Rådström P. Sample preparation methods in PCR-based detection of food pathogens. Trends Food Sci Technol. 1994; 5: 384-389. doi:10.1016/0924-2244(94)90166-X.

9. Border PM, Howard JJ, Plastow GS, Siggens KW. Detection of Listeria species and Listeria monocytogenes using polymerase chain reaction. Lett. Appl Microbiol.

Prevalence of Listeria Monocytogenes in Milk

1990; 11: 158–162. doi:https://doi.org/10.1111/j.1472-765X.1990.tb00149.x.

10. Arslan S, Özdemir F. Prevalence and antimicrobial resistance of Listeria spp. in homemade white cheese. Food Control. 2008; 19: 360–363. doi:https://doi.org/10.1016/j. foodcont.2007.04.009.

11. Rahimi E, Ameri M, Momtaz H. Prevalence and antimicrobial resistance of Listeria species isolated from milk and dairy products in Iran. Food Control. 2010; 21: 1448–1452. doi:https://doi.org/10.1016/j. foodcont.2010.03.014.

12. Pearson LJ, Marth EH. Listeria monocytogenes – Threat to a Safe Food Supply: A Review. J Dairy Sci. 1990; 73: 912–928. doi:10.3168/jds.S0022-0302(90)78748-6.

13. Durmaz H, Avci M, Aygun O. The presence of Listeria species in corn silage and raw milk produced in Southeast Region of Turkey. Kafkas Üniversitesi Vet Fakültesi Derg. 2105. 21: 41–44.

14. Beckers HJ, Soentoro PSS, Delgou-van Asch EHM. The occurrence of Listeria monocytogenes in soft cheeses and raw milk and its resistance to heat. Int J Food Microbiol. 1987; 4: 249–256. doi:10.1016/0168-1605(87)90041-9.

15. Derra FA, Karlsmose S, Monga DP, Mache A, Svendsen CA, Félix B, et al. Occurrence of Listeria spp. in Retail Meat and Dairy Products in the Area of Addis Ababa, Ethiopia. F Patho Dis. 2013; 10: 577–579. doi:10.1089/fpd.2012.1361.

16. Dalzini E, Bernini V, Bertasi B, Daminelli P, Losio M, Varisco G. Survey of prevalence and seasonal variability of Listeria monocytogenes in raw cow milk from Northern Italy. Food Control. 2016; 60: 466–470.

17. Cerva C, Bremm C, Marques dos Reis E, Vinícius Andrade Bezerra A, Regina Loiko M, Estêvão Farias da Cruz C, et al. Quoos Mayer F. Food safety in raw milk production: risk factors associated to bacterial DNA contamination. Trop Animal Health Prod. 2014; 46: 877–882. doi:10.1007/s11250-014-0580-y.

18. A. Abbas B, M. Jaber G. Occurrence of Listeria monocytogens in raw milk of ruminants. Iraqi J Vet Sci. 2012; 26: 47–51. doi:10.33899/ijvs.2012.46959.

19. Mahmoodi MM. Occurrence of Listeria monocytogenes in raw milk and dairy products in Noorabad, Iran. J Ani Vet Adv. 2010; 9: 16–19.

20. Bemrah N, Sanaa M, Cassin MH, Griffiths MW, Cerf O. Quantitative risk assessment of human listeriosis from consumption of soft cheese made from raw milk. Prev Vet Med. 1998; 37: 129–145. doi:10.1016/s0167-5877(98)00112-3.

21. Sagun E, Sancak YC, I_v'leyl_cl Ö, Ekici K. The Presence and Prevalence of Listeria Species in Milk and Herby Cheese in and Around Van. Turkish J Vet. Anim Sci. 2001; 25: 15–19.

22. Jamali H, Radmehr B, Thong KL. Prevalence, characterisation, and antimicrobial resistance of Listeria species and Listeria monocytogenes isolates from raw milk in farm bulk tanks. Food Control. 2013; 34: 121–125. doi:10.1016/j.foodcont.2013.04.023.

Ebadifar A, et al.

23. Akya A, Najafi F, Moradi J, Mohebi Z, Adabagher S. Prevalence of food contamination with Listeria spp. in Kermanshah, Islamic Republic of Iran. East. Mediterr. Health J. 2013; 19: 474–477.

24. D'Amico DJ, Donnelly CW. Detection, isolation, and incidence of Listeria spp. in small-scale artisan cheese processing facilities: A methods comparison. J. Food Prot. 2009; 72: 2499–2507.

25. Barancelli GV, Camargo TM, Reis CM, Porto E, Hofer E, Oliveira CA. Incidence of Listeria monocytogenes in cheese manufacturing plants from the northeast region of São Paulo, Brazil J Food Prot. 2011; 74: 816–819.

26. Nero LA, De Mattos MR, de Aguiar Ferreira Barros M, Ortolani MBT, Beloti V, de Melo Franco BDG. Listeria monocytogenes and Salmonella spp. in raw milk produced in Brazil: Occurrence and interference of indigenous microbiota in their isolation and development. Zoonoses Public Health. 2008; 55: 299–305.

27. Ortolani MBT, Yamazi AK, Moraes PM, Viçosa GN, Nero LA. Microbiological quality and safety of raw milk and soft cheese and detection of autochthonous lactic acid bacteria with antagonistic activity against Listeria monocytogenes, Salmonella spp. and Staphylococcus aureus. Foodborne Pathog. Dis. 2010; 7: 175–180.

28. Schoder D, Melzner D, Schmalwieser A, Zangana A, Winter P, Wagner M. Important vectors for Listeria monocytogenes transmission at farm dairies manufacturing fresh sheep and goat cheese from raw milk. J. Food Prot. 2011; 74: 919–924.

29. Ismaiel AAR, Ali AES, Enan G. Incidence of Listeria in Egyptian meat and dairy samples. Food Sci. Biotechnol. 2014; 23: 179–185. 30. Lopez CS, Sa'nchez H H. Behaviour of Listeria monocytogenes during the manufacture and ripening of Manchego and Chihuahua Mexican cheeses. Int J Food Microbiol. 2000; 62: 149–153. 31. Buyser ML, B. Dufour M, Maire V. Lafarge. Implication of milk and milk products in food-borne diseases in France and in different industrialised countries. Int. J. Food Microbiol. 2001; 67: 1–17.

32. Cordano AM, Rocourt J. Occurrence of Listeria monocytogenes in food in Chile. Int J Food Microbiol. 2001; 70: 175–178.

33. Dhanashree B, S. K. Otta I, Karunasagar W, Goebel, Karunasagar I. Incidence of Listeria spp. in clinical and food samples in Mangalore, India. Food Microbiol. 2003; 20: 447–453.

34.Jay J M, Loessner M J, Golden D A. Modern food microbiology. Ann. Microbiol. 2006; 56: 1- 81. https://doi.org/10.1007/BF03174975.

35. Pearson LJ, Marth EH. Listeria monocytogenes – Threat to a Safe Food Supply: A Review. J Dairy Sci. 1990; 73: 912–928. doi:10.3168/jds.S0022-0302(90)78748-6.

36. El-Gazzar FE, Bohner HF, Marth EH. Antagonism between Listeria monocytogenes and lactoccoci during fermentation of products from ultrfiltered skim milk. J Dairy Sci. 1992; 75(1): 43-50.

37. Theodoridis AK, Papagoregiou DK, Abrahim A, Karaioannogou PG. Fate of Listeria monocytogenes during the manufacture and storage of Chevre Metsovo and Pichtogalo Chanion cheeses (Greece). Italian J Food Sci. 2006; 18(1): 51-61.

38. Jayamanne V, Samarajeewa U. Evaluation of the heat resistance of pathogenic Listeria monocytogenes in milk and milk products in Sri Lanka. Trop Agric Res Ext. 2011; 13: 73–80.

39. Osman KM, Zolnikov TR, Samir A, Orabi A. Prevalence, pathogenic capability, virulence genes, biofilm formation, and antibiotic resistance of Listeria in goat and sheep milk confirms need of hygienic milking conditions. Pathog Glob Health. 2104; 108: 21–29. doi:10.1179/2047773213Y.0000000115.

[Downloaded from jabs.fums.ac.ir on 2025-08-31